



## Review

## Effect of stress on brain inflammation and multiple sclerosis

Anna Karagkouni <sup>a</sup>, Michail Alevizos <sup>a</sup>, Theoharis C. Theoharides <sup>a,b,c,d,e,\*</sup><sup>a</sup> Molecular Immunopharmacology and Drug Discovery Laboratory, Department of Molecular Physiology and Pharmacology, Tufts University School of Medicine, 136 Harrison Avenue, Boston, MA, USA<sup>b</sup> Department of Biochemistry, Tufts University School of Medicine, 136 Harrison Avenue, Boston, MA, USA<sup>c</sup> Department of Internal Medicine, Tufts University School of Medicine, 136 Harrison Avenue, Boston, MA, USA<sup>d</sup> Tufts Medical Center, 136 Harrison Avenue, Boston, MA, USA<sup>e</sup> Department of Psychiatry, Tufts University School of Medicine, 136 Harrison Avenue, Boston, MA, USA

## ARTICLE INFO

## Article history:

Received 18 February 2013

Accepted 28 February 2013

Available online 26 March 2013

## Keywords:

Blood–brain barrier

Corticotropin-releasing hormone

Flavonoids

Neurotensin

Neuropeptides

Inflammation

Microglia

Mast cells

Multiple sclerosis

Stress

## ABSTRACT

Substantial evidence indicates that stress can precipitate or worsen symptoms of inflammation in general and more specifically in multiple sclerosis (MS), a demyelinating, autoimmune disease characterized by inflammation of the central nervous system (CNS). However, the mechanism of how stress affects MS is not well understood. We reviewed publications in PubMed since 1995 and propose that neuropeptides secreted under stress, such as corticotropin releasing hormone (CRH) and neurotensin (NT), activate microglia and mast cells to release inflammatory molecules. These lead to maturation and activation of T17 autoimmune cells, disruption of the blood–brain barrier (BBB) and T cell entry into the CNS, thus promoting brain inflammation and contributing to MS pathology. Reduction of stress and inhibition of these processes by select flavonoids could provide novel therapeutic approaches.

© 2013 Elsevier B.V. All rights reserved.

## Contents

1. Introduction	947
2. Correlation between stress and MS	948
2.1. Human studies	948
2.2. Animal studies	948
3. Involvement of microglia and mast cells	949
4. Possible mechanisms to translate stress in MS risk	950
4.1. Stress and the HPA axis	950
4.2. Stress, microglia and mast cells	950
5. Conclusion	950
Authors' contributions	951
Disclosures	951
Conflicts	951
Take-home messages	951
References	951

\* Corresponding author at: Department of Molecular Physiology and Pharmacology, Tufts University School of Medicine, Suite J304, 136 Harrison Avenue, Boston, MA 02111, USA. Tel.: +1 617 636 6866; fax: +1 617 636 2456.

E-mail addresses: [anna.karagkouni@tufts.edu](mailto:anna.karagkouni@tufts.edu) (A. Karagkouni), [michail.alevizos@tufts.edu](mailto:michail.alevizos@tufts.edu) (M. Alevizos), [theoharis.theoharides@tufts.edu](mailto:theoharis.theoharides@tufts.edu) (T.C. Theoharides).

## 1. Introduction

Multiple sclerosis (MS) is an autoimmune disease of the central nervous system (CNS) characterized by brain inflammation, demyelination and axonal loss [1]. Clinically, the disease follows a relapsing–remitting or a progressive course. Many possible triggering factors

such as infections, toxins, immunizations, trauma, sunlight exposure and hormonal variables have been implicated [2]. Although MS affects young women more often than men, male gender has been associated with a poorer prognosis. This could be attributed to a possible protective role that estrogen or progesterone may have in the severity of the disease, but not in the risk of MS [3–5].

Increasing evidence indicates that stress can worsen immunity [6] and brain inflammation [7,8], which is important in the pathogenesis of MS [9], and neuropsychiatric disorders in general [10,11]. We reviewed publications in PubMed since 1995 that report any association between stress, neuropeptides, microglia, mast cells and MS.

## 2. Correlation between stress and MS

### 2.1. Human studies

Increasing evidence indicates that symptoms in relapsing–remitting MS may be precipitated or exacerbated by stress [12]. A meta-analysis of 14 studies (case–control and longitudinal prospective) published in 2004, showed that there is a significant association between stressful life events and subsequent relapses of MS in humans [13]. A cohort study in Denmark examined the association between MS and a major stressful life event, the death of a child. The study comprised 21,062 parents who had lost a child and 293,745 matched parents who had not lost a child. The study indicated a significantly higher MS risk in parents who lost a child than in parents who did not for at least 8 years [14]. MS relapses, in a population of 50 female MS patients in the USA, were more likely during at-risk periods following stressful life events and were relatively independent of the threat level and type of stressor [15]. In a study conducted in Greece, cumulative stressful life events were shown to pose a greater risk for relapse in ambulatory women with relapsing–remitting MS [16]. Duration was the only stress attribute that seemed to increase the risk for relapsing in contrast to stress type and stress severity [16]. In another Greek study, 37 female patients kept diaries of stressful life events and anxiety levels that were subsequently ranked according to the Holmes and Rahe Social Readjustment Rating Scale and the Hamilton Rating Scale for Anxiety, respectively. Multiple reported stressful life events and elevated levels of anxiety were each found to be significantly associated with increased risk for relapse of MS [17]. A Lebanese study of 216 patients showed an increase in both clinical relapses and MRI disease activity in patients with MS during periods of war stress [18]. This is in line with an earlier study of 156 patients in Israel that reported that civilian exposure to war stress is associated with increased risk for MS relapses [19].

In a case–control study of 100 MS patients compared to hospital controls, significantly more MS patients reported that they were under unusual stress in the 2 year period prior to onset age [20]. Also, a comparison of 95 pairs of MS patients revealed that patients in relapse scored higher on emotional disturbance and intensity of stressful events than patients in remission [21]. An American study of 55 MS patients showed that patients who experienced qualitatively extreme stressful events were 37 times more likely to relapse than those not exposed to such events [22]. A comparison of 39 patients with early MS and 40 matched non-patient volunteers in another American study revealed that the proportion of MS patients who experienced marked life adversity in the year prior to onset of symptoms was significantly higher than for non-patients in the year before interview [23]. Similar results were reported in a study of 73 patients in Netherlands in which stressful events were associated with increased relapses in relapsing–remitting MS [24]. Finally, in a one year study, 48 women with relapsing–remitting MS were divided into two groups, either receiving the anti-anxiety/anti-depressant drug escitalopram daily or continuing with MS treatment as usual, and stressful life events were documented weekly. The risk for relapse was 2.9 times higher for controls than for the escitalopram-treated patients [25]. Table 1 lists studies reporting stress–MS association in humans.

**Table 1**  
Studies in humans reporting correlation between stress and MS.

Type of stress	n	Outcome	Reference
Death of child	21,062 parents exposed 293,745 not exposed	The exposed parents had a significant increased risk of MS for at least 8 years of follow-up.	[14]
Stressful life events (SLE) as reported weekly assessed with the Life Events and Difficulties Schedule	50	Relapses were more likely during at-risk periods following the events.	[15]
Diaries of SLE ranked according to the Holmes and Rahe Social Readjustment Rating Scale. Anxiety assessed with the Hamilton Rating Scale for anxiety	37	High levels of anxiety were strongly related with the advent of relapse in the following period.	[17]
War stress	216	Significant increase in the number of relapses during war period than during non-war periods	[18]
War stress	156	Increased number of relapses during the war period	[19]
SLE in self-report diaries classified as short or long-term. Severity determined using the Recent Life Change Questionnaire	26	Three or more SLE in a 4 week period led to 5 times increase of MS relapse rate.	[16]
Life stress	100 MS patients compared to controls	Significantly more MS patients than controls reported that they were under unusual stress in the 2 year period prior to onset age. MS patients described a greater number of SLE.	[20]
SLE assessed by the Psychiatric Epidemiology Research Interview	55	Patients who experienced qualitatively extreme events were 37 times more likely to relapse as those not exposed to such events.	[22]
SLE	39 MS patients and 40 matched controls	The proportion of MS patients who experienced marked life adversity in the year prior to onset of symptoms was significantly higher than for non-patients in the year before interview.	[23]
Emotional stress	95 pairs of MS patients in relapse and remission	Patients with MS exacerbation scored higher on emotional disturbance and intensity of stressful events than patients in remission.	[21]

### 2.2. Animal studies

In mice with Theiler's murine encephalomyelitis virus (TMEV) infection, a well characterized model of MS, restraint stress during early infection increased CNS lesion formation during the late phase [26]. Restraint stress in animals infected with TMEV had a global

immunosuppressive effect on the immune response to infection [27], leading to increased mortality rates, decreased numbers of lymphocytes and increased numbers of neutrophils in the blood [28]. This paradigm also exacerbated acute CNS infection and subsequent demyelination [29]. Maternal separation (180 min/day) impaired host resistance during infection and prolonged TMEV [30]. A significant increase in the severity of neurological signs was noted along with pathological lesions of the spinal cord in stressed rats compared to non-stressed rats; treatment with alprazolam reversed the adverse effects of stress [31]. However, another study using Theiler's virus-induced demyelinating disease (TVID) in the resistant C57BL/6 mouse strain suggested that stress alone is not sufficient to overcome genetic resistance to TVID [32]. Acute restraint stress disrupted the BBB and substituted for diphtheria toxin permitting the development of myelin basic protein-induced EAE sooner in rats [33]. Administration of diazepam for 6 days, starting at day 6 or 11 after active induction of experimental allergic encephalomyelitis (EAE), an animal model for MS, led to a marked decrease of disease incidence, reduced histological signs associated with the disease, as well as cellular reactivity and antibody responses against the encephalitogenic MBP [34]. Table 2 lists studies showing stress–EAE association in animals.

### 3. Involvement of microglia and mast cells

Recent evidence indicates that microglia play an important role in the pathogenesis of MS. The most intense microglia infiltration has been observed in acute MS cases in which the acute stage inflammatory macrophage markers MRP14 and 27E10 were expressed [35]. It has also been shown that in a specific subtype of MS, where hypoxia-like lesions exist, microglial activation is prominent and precedes T-cell infiltration and demyelination [36]. Moreover, brain pathological findings from patients who died of MS exhibited extensive oligodendrocyte apoptosis and microglial activation in the relative absence of T-cells [37]. Actually,

microglia act as antigen-presenting cells for naïve T-cells, thus expanding the number of encephalitogenic Th1 cells [38]. Moreover, microglia have the ability to produce glutamate and nitric oxide (NO), which have a direct effect on the death of neurons. NO also has a cytotoxic effect on the endothelium and contributes to the BBB disruption [38], which is known to precede many pathological or clinical symptoms of MS [39,40]. Furthermore, dying oligodendroglial cells recruit microglia which, in the presence of IFN- $\gamma$  activation, induce contact-dependent oligodendroglial death [41]. Lastly, microglia are a rich source of reactive oxygen species (ROS), and various pro-inflammatory cytokines/chemokines and proteases [42].

On the other hand, microglia might have a role in the termination of the inflammatory reaction by suppressing lymphocyte reactivity through NO release [43]. In fact, a strong accumulation of CD 163(+) microglia with anti-inflammatory effects was found in acute active MS lesions and at the rim of chronic active lesions, possibly involved in the resolution of the inflammation [44]. Microglia also phagocytose apoptotic T-cells, even though this mechanism seems to be defective in MS [38].

It has been shown that in mice with EAE, microglial activation persists during the chronic phase of the disease, while T cell infiltrates are predominant during the acute phase of the disease [45]. Microglia participate in the pathogenesis of EAE not only by phagocytosing myelin and thus leading to demyelination [46], but also by releasing TNF- $\alpha$ , IL-1, IL-6 and chemokines, which promote inflammation during the course of the disease [47]. In fact, Lewis rats which are susceptible to EAE showed suppression of disease progression upon elimination of microglia [48].

MS is mediated primarily by brain infiltration of Th1 cells and macrophages [49], but Th2 processes typically associated with allergic reactions, which involve mast cells, are also implicated [50–52]. Mast cells have been reported in MS plaques [53] and could stimulate demyelination directly [54–56]. Clinical evidence supporting the involvement of brain mast cells in MS comes from the fact that the unique mast cell protease tryptase [57] and histamine [58] were elevated in the CSF of MS patients. Moreover, gene microarray analysis of MS plaques revealed increased expression of 5-lipoxygenase in acute lesions and the Fc $\epsilon$ R1 receptor in chronic lesions, both of which are associated with mast cells [59,60]. Mast cells are also involved in Th17 maturation. Th17 cells are differentiated by the combined action of IL-6 and TGF- $\beta$  to secrete IL-17 [61], shown to be critical for the pathogenesis of autoimmune diseases, including MS [61,62]. TNF- $\alpha$  and vasoactive intestinal peptide (VIP) can also induce Th17 maturation independently of IL-6 [63]. It is noteworthy that IL-6, TGF- $\beta$ , TNF- $\alpha$ , and VIP can all be secreted by mast cells [64–66]. In fact, mast cells can even secrete IL-17 on their own [67]. Mast cell mediators can recruit and activate T cells, as well as permit them to enter the brain by disrupting the BBB [68]. Mast cells stimulated by Fc $\epsilon$ R1 aggregation released TNF- $\alpha$  and activated T cells [69], but direct contact was also required [70]. Mast cell-derived leukotriene B4 promotes T cell migration [71]. Fig. 1 proposes a set of interactions between mast cells and T-cells.

In EAE, mast cells are required for optimal T cell responses [72], but can also degrade myelin directly [54,56]. Development of EAE had been shown to involve mast cell accumulation in the rat brain [73] that could be due to chemotactic activity elicited by RANTES [74] or MCP-1 [75] secreted from either glial cells or infiltrating leukocytes. EAE was attenuated and delayed in W/W<sup>v</sup> mast-cell deficient mice [76], but was fully restored upon mast cell reconstitution even in the apparent absence of brain mast cell replenishment [77]. This effect apparently required mast cells outside the brain [78], especially in the meninges [79]. The authors concluded that brain mast cells are not important, but did not exclude the involvement of perivascular mast cells and their ability to regulate the permeability of BBB [80,81].

Interestingly, some recent data appear to suggest that mast cells may also have a protective effect on EAE development. Specifically, both activating and suppressing Fc receptors were recently shown to be expressed on mast cells and regulate EAE disease severity in

**Table 2**  
Studies in animals reporting correlation between stress and EAE.

Type of stress	Outcome	Reference
Restraint stress (mice restrained in ventilated 60 ml plastic syringes in their home cages)	Inflammation and demyelination were significantly increased in spinal cords of stressed mice. Axonal degradation was increased in demyelinated areas in stressed mice.	[26]
Restraint stress	Stress had a global immunosuppressive effect on the immune response to infection. The adverse effects of stress were mimicked by dexamethasone, implicating a major role for glucocorticoids.	[27]
Restraint stress (mice placed in well ventilated restraining tubes, 2–3 cm internal diameter and 8 cm length)	Stress decreased both type 1 and type 2 responses to infection.	[141]
Maternal separation	Maternal separation 180 min/day impaired host resistance during infection and delayed the kinetics of viral clearance.	[30]
Restraint stress (mice placed in well ventilated restraining tubes, 2–3 cm internal diameter and 8 cm length)	Increased mortality rates were observed in restrained mice, which also developed higher CNS viral titers. Restrain-stressed mice developed decreased numbers of lymphocytes and increased numbers of neutrophils in the blood.	[28]
Social disruption stress	Social disruption stress applied prior to infection led to more severe disease course, with increased inflammation.	[93]
Restraint stress (mice restrained in ventilated 60 ml plastic syringes in their home cages)	Chronic restraint stress during early infection exacerbated acute CNS infection and the subsequent demyelination.	[29]

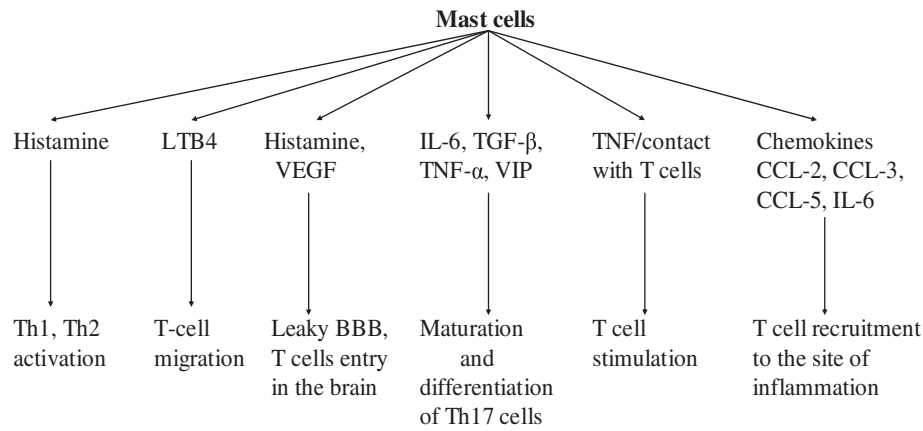


Fig. 1. Diagrammatic representation of the proposed interactions of mast cells with T-cells.

mice [82]. Myelin oligodendrocyte glycoprotein (MOG)<sub>35–55</sub>-induced EAE was exacerbated in mast-cell deficient  $Kit^{W-sh/W-sh}$  mice both at high and low antigen protocols, while  $Kit^{W/W-v}$  mice were protected when immunized with high, but not low doses of antigen [83]. In addition, when the mast-cell deficient mice were reconstituted with bone marrow derived mast cells, systemically, but not in the CNS, they still developed exacerbated EAE [83]. Any protective role against EAE may be exerted by Treg activation only in the brain. In fact, EAE in  $Kit^{W-sh/W-sh}$  mice developed earlier with more severe clinical and pathological symptoms; reconstitution with mast cells reduced susceptibility to the disease, and correlated with mast cell recruitment and  $T_{reg}$  activation in the CNS [84]. Yet, mast cell-derived TNF exacerbated mortality during severe infection that was reduced in  $Kit^{W/W-v}$  mast-cell deficient mice [85]. Moreover, EAE development was reduced in TNF knock-out mice [86], but became worse at a later date [86]. It is quite critical that results obtained from mouse models should be interpreted cautiously as they may not be readily applicable to humans [87].

#### 4. Possible mechanisms to translate stress in MS risk

##### 4.1. Stress and the HPA axis

Stress activates the hypothalamic–pituitary–adrenal (HPA) axis through the hypothalamic secretion of corticotropin-releasing hormone (CRH), which normally suppresses immune responses through the release of glucocorticoids from the adrenals [88]. In this context, it has been proposed that MS worsening with stress may be due to dysfunctional HPA axis because of reduced production of adrenal steroids. Impaired activation of CRH neurons was shown in those with active MS lesions in the hypothalamus, where the disease course was more severe [89]. Patients with secondary progressive MS also exhibit less cortisol production in response to CRH stimulation [90]. The absence of a normal cortisol response during systemic infection was reported in patients with MS suggesting impaired cortisol secretion and a reduced ability to control inflammation [91]. Despite the fact that clinical conditions characterized by overproduction of pro-inflammatory cytokines are associated with elevated circulating cortisol levels, this is not observed in MS patients [92]. Development of glucocorticoid resistance due to stress exposure may also result in increased CNS inflammation [93].

A temporal framework has also been proposed in order to explain the effects of stressful life events in patients with MS [94]. Specifically, acute stress might have a permissive effect on MS exacerbation by facilitating BBB breakdown, while chronic stress may lead to glucocorticoid resistance, making the immune cells less responsive to regulatory control by cortisol.

##### 4.2. Stress, microglia and mast cells

CRH also has pro-inflammatory effects [95,96]. CRH affects brain microvessels directly [97] and activates mast cells [97,98] leading to increased BBB permeability which was absent in mast-cell deficient mice [99,98]. We have also reported that CRH and NT which are secreted under stress, synergistically stimulate mast cells leading to increased vascular permeability [100] and blood–brain barrier (BBB) disruption [101]. We further showed that NT stimulates mast cell secretion of vascular endothelial growth factor (VEGF) [102], which increases BBB permeability. NT also induces expression of CRH receptor-1 (CRHR-1) [103], activation of which by CRH increases the stimulation of human mast cells [104]. Animal experiments showed that acute stress led to BBB disruption in rats [105,106] and shortened the time of onset of EAE [33]. In fact, EAE could not develop in CRH knockout mice [107].

Stress activates microglia as well [108–110]. Specifically, exposure of rats to cold stress provoked morphological activation of microglia [108]. In addition, restraint stress combined with water immersion stress induced morphological activation of microglia in the thalamus, hypothalamus, hippocampus, substantia nigra, central gray, an effect that was significantly reduced in rats null for IL-18 [109].

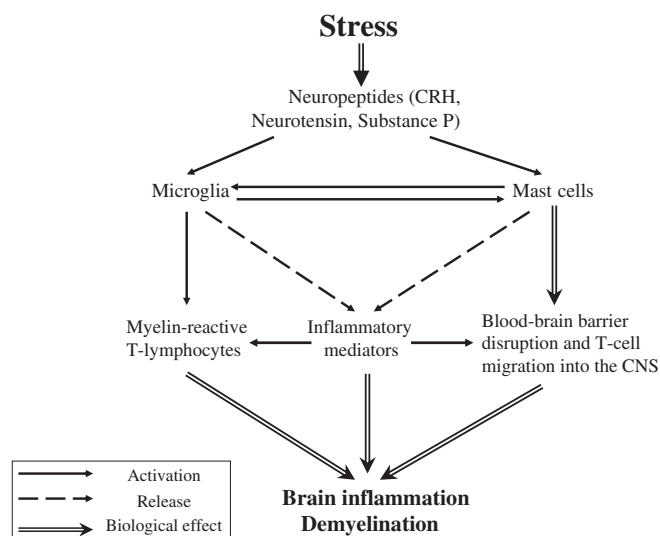
CRH induces the proliferation and TNF- $\alpha$  release by cultured rat microglial cells [111]. Microglia also express neurotensin (NT) receptor 3 (NTR3) leading to their proliferation and gene expression of macrophage inflammatory protein-2 (MIP-2), MCP-1, interleukin-1beta and TNF- $\alpha$ . [112]. SP receptors have been detected in both murine and human microglia, activation of which by SP led to the activation of NF- $\kappa$ B transcriptional factor [42].

Human microglia have also been shown to produce SP [42], which is known to activate mast cells [113]. On the other hand, microglia respond to pro-inflammatory signals released from mast cells [114]. Mast cell tryptase induces microglial activation and pro-inflammatory mediator release of TNF- $\alpha$ , IL-6 and ROS [115]. Emerging evidence suggests that mast cell–microglial interactions play an important role in neuroinflammatory diseases [114]. Fig. 2 shows possible interactions between stress, microglia, mast cells and brain inflammation.

#### 5. Conclusion

There have been important new treatment options for MS patients [116]. Moreover, recent evidence indicates that glatiramer acetate, in addition to having an immunomodulatory [117] and a neuroprotective effect [118], also decreases TNF- $\alpha$  while increasing IL-10 secretion and promoting phagocytic activity of microglia [119].

However, addressing the effect of stress on MS is an entirely new treatment option. For instance, in a study of 121 patients with relapsing



**Fig. 2.** Diagrammatic representation of the effect of stress on brain inflammation and the proposed interactions between microglia and mast cells.

MS, stress management therapy resulted in the reduction of the development of new MRI lesions [120]. Additionally, in a study of 62 patients with MS, it was shown that those who attended an 8-week stress-management program experienced a decreased number of weekly symptoms and mean intensity per symptom [121]. Moreover, publications using animal models reported that diazepam [34] or alprazolam [31] can decrease or reverse the clinical and histological signs associated with EAE. CRH antagonists [122] may also be useful since EAE could not develop in CRH knockout mice [107].

Microglia [36] and mast cells [80,123] have been considered as the next therapeutic targets for MS. However, there are no clinically available inhibitors of these cell types. Certain natural flavonoids, such as quercetin, luteolin and apigenin have anti-oxidant and anti-inflammatory effects [124]. These flavonoids also suppress TNF- $\alpha$  and IL-6 expression and release from microglia [125–127], as well as mast cell activation [128] and release of cytokines [129–131]. The flavonoids luteolin and quercetin decrease the amount of myelin phagocytosed by macrophages [132], as well as reduce EAE [133–135]. Luteolin also inhibits mast cell-dependent T cell activation [136]. Apigenin sensitizes activated human T cells to apoptosis and inhibits auto-antigen-presenting cells necessary for the expansion and activation of Th17 cells in lupus [137]. Propolis, a flavonoid-containing substance, inhibits IL-6 plus TGF- $\beta$ -induced Th17 differentiation in vitro [138]. Luteolin inhibits activated peripheral blood mononuclear cells and had synergistic effect with IFN- $\beta$  [139,140] prompting the suggestion that luteolin may be a reasonable adjuvant for MS treatment [139].

### Authors' contributions

All authors have read and approved the final manuscript. TCT designed and wrote most of the paper. AK and MA researched the literature and prepared the manuscript.

### Disclosures

Dr. Theoharides is the inventor of US Patents No. 7906153 covering the use of flavonoids in the treatment of MS and No. 8268365 covering the use of flavonoids in the treatment of brain inflammation.

### Conflicts

The authors report no conflict of interest.

### Take-home messages

- Human and animal studies show a correlation between stress and relapses of multiple sclerosis.
- Microglia and mast cells are involved in the pathogenesis of multiple sclerosis.
- Stress activates microglia and mast cells through the release of the neuropeptide neurotensin and corticotropin-releasing hormone.
- Stress-induced activation of microglia and mast cells leads to BBB disruption and brain inflammation.
- Stress reduction as well as inhibition of microglial and mast cell activation can prove to be a useful adjunct to the current treatments of multiple sclerosis.

### References

- [1] Bruck W. The pathology of multiple sclerosis is the result of focal inflammatory demyelination with axonal damage. *J Neurol* 2005;252(Suppl. 5):v3–9.
- [2] Casetta I, Granieri E. Prognosis of multiple sclerosis: environmental factors. *Neurol Sci* 2000;21:S839–42.
- [3] Nussinovitch U, Shoenfeld Y. The role of gender and organ specific autoimmunity. *Autoimmun Rev* 2012;11:A377–85.
- [4] Lee TP, Chiang BL. Sex differences in spontaneous versus induced animal models of autoimmunity. *Autoimmun Rev* 2012;11:A422–9.
- [5] Hughes GC. Progesterone and autoimmune disease. *Autoimmun Rev* 2012;11:A502–14.
- [6] Nakata A. Psychosocial job stress and immunity: a systematic review. *Methods Mol Biol* 2012;934:39–75.
- [7] Angelidou A, Asadi S, Alysandratos KD, Karagkouni A, Kourembanas S, Theoharides TC. Perinatal stress, brain inflammation and risk of autism – review and proposal. *BMC Pediatr* 2–7-2012;12:89.
- [8] Garate I, Garcia-Bueno B, Madrigal JL, Caso JR, Alou L, Gomez-Lus ML, et al. Stress-induced neuroinflammation: role of the Toll-like receptor-4 pathway. *Biol Psychiatry* 1-1-2013;73:32–43.
- [9] Frischer JM, Bramow S, Dal-Bianco A, Lucchinetti CF, Rauschka H, Schmidbauer M, et al. The relation between inflammation and neurodegeneration in multiple sclerosis brains. *Brain* 2009;132:1175–89.
- [10] Theoharides TC, Zhang B, Conti P. Decreased mitochondrial function and increased brain inflammation in bipolar disorder and other neuropsychiatric diseases. *J Clin Psychopharmacol* 2011;31:685–7.
- [11] Hagberg H, Gressens P, Mallard C. Inflammation during fetal and neonatal life: implications for neurologic and neuropsychiatric disease in children and adults. *Ann Neurol* 2012;71:444–57.
- [12] Artemiadis AK, Anagnostouli MC, Alexopoulos EC. Stress as a risk factor for multiple sclerosis onset or relapse: a systematic review. *Neuroepidemiology* 2011;36:109–20.
- [13] Mohr DC, Hart SL, Julian L, Cox D, Pelletier D. Association between stressful life events and exacerbation in multiple sclerosis: a meta-analysis. *BMJ* 2004;328:731.
- [14] Li J, Johansen C, Bronnum-Hansen H, Stenager E, Koch-Henriksen N, Olsen J. The risk of multiple sclerosis in bereaved parents: a nationwide cohort study in Denmark. *Neurology* 2004;62:726–9.
- [15] Ackerman KD, Stover A, Heyman R, Anderson BP, Houck PR, Frank E, et al. 2002 Robert Ader New Investigator award. Relationship of cardiovascular reactivity, stressful life events, and multiple sclerosis disease activity. *Brain Behav Immun* 2003;17:141–51.
- [16] Mitsionis CI, Zervas IM, Mitropoulos PA, Dimopoulos NP, Soldatos CR, Potagas CM, et al. The impact of stressful life events on risk of relapse in women with multiple sclerosis: a prospective study. *Eur Psychiatry* 2008;23:497–504.
- [17] Potagas C, Mitsionis C, Watier L, Dellatolas G, Retziou A, Mitropoulos P, et al. Influence of anxiety and reported stressful life events on relapses in multiple sclerosis: a prospective study. *Mult Scler* 2008;14:1262–8.
- [18] Yamout B, Itani S, Hourany R, Sibaii AM, Yaghi S. The effect of war stress on multiple sclerosis exacerbations and radiological disease activity. *J Neurol Sci* 15-1-2010;288:42–4.
- [19] Golan D, Somer E, Dishon S, Cuzin-Disegni L, Miller A. Impact of exposure to war stress on exacerbations of multiple sclerosis. *Ann Neurol* 2008;64:143–8.
- [20] Warren S, Greenhill S, Warren KG. Emotional stress and the development of multiple sclerosis: case control evidence of a relationship. *J Chronic Dis* 1982;35:821–31.
- [21] Warren S, Warren KG, Cockerill R. Emotional stress and coping in multiple sclerosis (MS) exacerbations. *J Psychosom Res* 1991;35:37–47.
- [22] Franklin GM, Nelson LM, Heaton RK, Burks JS, Thompson DS. Stress and its relationship to acute exacerbations in multiple sclerosis. *J Neurol Rehabil* 1988;2:7–11.
- [23] Grant I, Brown GW, Harris T, McDonald WI, Patterson T, Trimble MR. Severely threatening events and marked life difficulties preceding onset or exacerbation of multiple sclerosis. *J Neurol Neurosurg Psychiatry* 1989;52:8–13.
- [24] Buljevac D, Hop WC, Reedeker W, Janssens AC, van der Meche FG, van Doorn P, et al. Self reported stressful life events and exacerbations in multiple sclerosis: prospective study. *BMJ* 2003;327:646.

- [25] Mitsonis CI, Zervas IM, Potagas CM, Mitropoulos PA, Dimopoulos NP, Sfagos CA, et al. Effects of escitalopram on stress-related relapses in women with multiple sclerosis: an open-label, randomized, controlled, one-year follow-up study. *Eur Neuropsychopharmacol* 2010;20:123–31.
- [26] Young EE, Sieve AN, Vichaya EG, Carcoba LM, Young CR, Ambrus A, et al. Chronic restraint stress during early Theiler's virus infection exacerbates the subsequent demyelinating disease in SJL mice: II. CNS disease severity. *J Neuroimmunol* 30-3-2010;220:79–89.
- [27] Welsh CJ, Steelman AJ, Mi W, Young CR, Dean DD, Storts R, et al. Effects of stress on the immune response to Theiler's virus – implications for virus-induced autoimmunity. *Neuroimmunomodulation* 2010;17:169–72.
- [28] Campbell T, Meagher MW, Sieve A, Scott B, Storts R, Welsh TH, et al. The effects of restraint stress on the neuropathogenesis of Theiler's virus infection: I. Acute disease. *Brain Behav Immun* 2001;15:235–54.
- [29] Sieve AN, Steelman AJ, Young CR, Storts R, Welsh TH, Welsh CJ, et al. Chronic restraint stress during early Theiler's virus infection exacerbates the subsequent demyelinating disease in SJL mice. *J Neuroimmunol* 2004;155:103–18.
- [30] Meagher MW, Sieve AN, Johnson RR, Satterlee D, Belyavskiy M, Mi W, et al. Neonatal maternal separation alters immune, endocrine, and behavioral responses to acute Theiler's virus infection in adult mice. *Behav Genet* 2010;40:233–49.
- [31] Núñez-Iglesias MJ, Novio S, Almeida-Dias A, Freire-Garabal M. Inhibitory effects of alprazolam on the development of acute experimental autoimmune encephalomyelitis in stressed rats. *Pharmacol Biochem Behav* 2010;97:350–6.
- [32] Steelman AJ, Alford E, Young CR, Welsh TH, Meagher MW, Welsh CJ. Restraint stress fails to render C57BL/6 mice susceptible to Theiler's virus-induced demyelination. *Neuroimmunomodulation* 2010;17:109–19.
- [33] Chandler N, Jacobson S, Connolly R, Esposito P, Theoharides TC. Acute stress shortens the time of onset of experimental allergic encephalomyelitis (EAE) in SJL/J mice. *Brain Behav Immun* 2002;16:757–63.
- [34] Bibolini MJ, Chanaday NL, Baez NS, Degano AL, Monferran CG, Roth GA. Inhibitory role of diazepam on autoimmune inflammation in rats with experimental autoimmune encephalomyelitis. *Neuroscience* 29-12-2011;199:421–8.
- [35] Bruck W, Sommermeier N, Bergmann M, Zettl U, Goebel HH, Kretzschmar HA, et al. Macrophages in multiple sclerosis. *Immunobiology* 1996;195:588–600.
- [36] Friese MA, Fugger L. T cells and microglia as drivers of multiple sclerosis pathology. *Brain* 2007;130:2755–7.
- [37] Barnett MH, Prineas JW. Relapsing and remitting multiple sclerosis: pathology of the newly forming lesion. *Ann Neurol* 2004;55:458–68.
- [38] Sanders P, De KJ. Janus faces of microglia in multiple sclerosis. *Brain Res Rev* 2007;54:274–85.
- [39] De Vreis HE, Kuiper J, de Boer AG, Van Berkel TJC, Breimer DD. The blood–brain barrier in neuroinflammatory diseases. *Pharmacol Rev* 1997;49:143–55.
- [40] Kwon EE, Prineas JW. Blood–brain barrier abnormalities in longstanding multiple sclerosis lesions. An immunohistochemical study. *J Neuropathol Exp Neurol* 1994;53:625–36.
- [41] Nicholas R, Stevens S, Wing M, Compston A. Oligodendroglial-derived stress signals recruit microglia in vitro. *NeuroReport* 23-5-2003;14:1001–5.
- [42] Kettenmann H, Hanisch UK, Noda M, Verkhratsky A. Physiology of microglia. *Physiol Rev* 2011;91:461–553.
- [43] Bogie JF, Stinissen P, Hellings N, Hendriks JJ. Myelin-phagocytosing macrophages modulate autoreactive T cell proliferation. *J Neuroinflammation* 2011;8:85.
- [44] Zhang Z, Zhang ZY, Schittenhelm J, Wu Y, Meyermann R, Schluessener HJ. Parenchymal accumulation of CD163+ macrophages/microglia in multiple sclerosis brains. *J Neuroimmunol* 15-8-2011;237:73–9.
- [45] Rasmussen S, Wang Y, Kivisakk P, Bronson RT, Meyer M, Imitola J, et al. Persistent activation of microglia is associated with neuronal dysfunction of callosal projecting pathways and multiple sclerosis-like lesions in relapsing–remitting experimental autoimmune encephalomyelitis. *Brain* 2007;130:2816–29.
- [46] Bauer J, Sminia T, Wouterlood FG, Dijkstra CD. Phagocytic activity of macrophages and microglial cells during the course of acute and chronic relapsing experimental autoimmune encephalomyelitis. *J Neurosci Res* 1-7-1994;38:365–75.
- [47] Benveniste EN. Role of macrophages/microglia in multiple sclerosis and experimental allergic encephalomyelitis. *J Mol Med* 1997;75:165–73.
- [48] Huitinga I, van RN, De Groot CJ, Uitdehaag BM, Dijkstra CD, Exp Med J. Suppression of experimental allergic encephalomyelitis in Lewis rats after elimination of macrophages. *J Exp Med* 1-10-1990;172:1025–33.
- [49] Batoulis H, Addicks K, Kuerten S. Emerging concepts in autoimmune encephalomyelitis beyond the CD4/T(H)1 paradigm. *Ann Anat* 20-8-2010;192:179–93.
- [50] Lassmann H, Ransohoff RM. The CD4–Th1 model for multiple sclerosis: a crucial re-appraisal. *Trends Immunol* 2004;25:132–7.
- [51] Pedotti R, De Voss JJ, Steinman L, Galli SJ. Involvement of both 'allergic' and 'autoimmune' mechanisms in EAE, MS and other autoimmune diseases. *Trends Immunol* 2003;24:479–84.
- [52] Pedotti R, DeVoss JJ, Youssef S, Mitchell D, Wedemeyer J, Madanat R, et al. Multiple elements of the allergic arm of the immune response modulate autoimmune demyelination. *Proc Natl Acad Sci U S A* 2003;100:1867–72.
- [53] Krüger PG, Bo L, Myhr KM, Karlens AE, Taule A, Nyland HI, et al. Mast cells and multiple sclerosis: a light and electron microscopic study of mast cells in multiple sclerosis emphasizing staining procedures. *Acta Neurol Scand* 1990;81:31–6.
- [54] Johnson D, Seelldrayers PA, Weiner HL. The role of mast cells in demyelination. 1. Myelin proteins are degraded by mast cell proteases and myelin basic protein and P<sub>2</sub> can stimulate mast cell degranulation. *Brain Res* 1988;444:195–8.
- [55] Brenner T, Soffer D, Shalit M, Levi-Schaffer F. Mast cells in experimental allergic encephalomyelitis: characterization, distribution in the CNS and in vitro activation by myelin basic protein and neuropeptides. *J Neurol Sci* 1994;122:210–3.
- [56] Theoharides TC, Dimitriadou V, Letourneau RJ, Rozniecki JJ, Vliagoftis H, Boucher WS. Synergistic action of estradiol and myelin basic protein on mast cell secretion and brain demyelination: changes resembling early stages of demyelination. *Neuroscience* 1993;57:861–71.
- [57] Rozniecki JJ, Hauser SL, Stein M, Lincoln R, Theoharides TC. Elevated mast cell tryptase in cerebrospinal fluid of multiple sclerosis patients. *Ann Neurol* 1995;37:63–6.
- [58] Tuomisto L, Kilpeläinen H, Riekkinen P. Histamine and histamine-N-methyltransferase in the CSF of patients with multiple sclerosis. *Agents Actions* 1983;13:255–7.
- [59] Tompkins SM, Miller SD. An array of possibilities for multiple sclerosis. *Nat Med* 2002;8:451–3.
- [60] Lock C, Hermans G, Pedotti R, Brendolan A, Schadt E, Garren H, et al. Gene-microarray analysis of multiple sclerosis lesions yields new targets validated in autoimmune encephalomyelitis. *Nat Med* 2002;8:500–8.
- [61] Weaver CT, Hattori RD, Mangan PR, Harrington LE. IL-17 family cytokines and the expanding diversity of effector T cell lineages. *Annu Rev Immunol* 2007;25:821–52.
- [62] Weaver CT, Harrington LE, Mangan PR, Gavrieli M, Murphy KM. Th17: an effector CD4 T cell lineage with regulatory T cell ties. *Immunity* 2006;24:677–88.
- [63] Kimura A, Naka T, Kishimoto T. IL-6-dependent and -independent pathways in the development of interleukin 17-producing T helper cells. *Proc Natl Acad Sci U S A* 2007;104:12099–104.
- [64] Theoharides TC, Alysandratos KD, Angelidou A, Delivanis DA, Sismanopoulos N, Zhang B, et al. Mast cells and inflammation. *Biochim Biophys Acta* 23-12-2010;1822:21–33.
- [65] Hugle T, Hogan V, White KE, van Laar JM. Mast cells are a source of transforming growth factor beta in systemic sclerosis. *Arthritis Rheum* 2011;63:795–9.
- [66] Gordon JR, Galli SJ. Mast cells as a source of both preformed and immunologically inducible TNF- $\alpha$ /cachectin. *Nature* 1990;346:274–6.
- [67] Lin AM, Rubin CJ, Khandpur R, Wang JY, Riblett M, Yalavarthi S, et al. Mast cells and neutrophils release IL-17 through extracellular trap formation in psoriasis. *J Immunol* 1-7-2011;187:490–500.
- [68] Theoharides TC, Kempuraj D, Kourelis T, Manola A. Human mast cells stimulate activated T cells: implications for multiple sclerosis. *Ann N Y Acad Sci* 2008;1144:74–82.
- [69] Nakae S, Suto H, Kakurai M, Sedgwick JD, Tsai M, Galli SJ. Mast cells enhance T cell activation: Importance of mast cell-derived TNF. *Proc Natl Acad Sci U S A* 3-5-2005;102:6467–72.
- [70] Salamon P, Shoham NG, Gavrieli R, Wolach B, Mekori YA. Human mast cells release interleukin-8 and induce neutrophil chemotaxis on contact with activated T cells. *Allergy* 2005;60:1316–9.
- [71] Ott VL, Cambier JC, Kappler J, Marrack P, Swanson BJ. Mast cell-dependent migration of effector CD8+ T cells through production of leukotriene B<sub>4</sub>. *Nat Immunol* 2003;4:974–81.
- [72] Gregory GD, Robbie-Ryan M, Secor VH, Sabatino Jr JJ, Brown MA. Mast cells are required for optimal autoreactive T cell responses in a murine model of multiple sclerosis. *Eur J Immunol* 2005;35:3478–86.
- [73] Dimitriadou V, Pang X, Theoharides TC. Hydroxyzine inhibits experimental allergic encephalomyelitis (EAE) and associated brain mast cell activation. *Int J Immunopharmacol* 2000;22:673–84.
- [74] Conti P, Reale M, Barbacane RC, Letourneau R, Theoharides TC. Intramuscular injection of hrRANTES causes mast cell recruitment and increased transcription of histidine decarboxylase: lack of effects in genetically mast cell-deficient W/W<sup>v</sup> mice. *FASEB J* 1998;12:1693–700.
- [75] Conti P, Pang X, Boucher W, Letourneau R, Reale M, Barbacane RC, et al. Impact of Rantes and MCP-1 chemokines on in vivo basophilic mast cell recruitment in rat skin injection model and their role in modifying the protein and mRNA levels for histidine decarboxylase. *Blood* 1997;89:4120–7.
- [76] Secor VH, Secor WE, Gutekunst C-A, Brown MA. Mast cells are essential for early onset and severe disease in a murine model of multiple sclerosis. *J Exp Med* 2000;191:813–21.
- [77] Brown MA, Tanzola M, Robbie-Ryan M. Mechanisms underlying mast cell influence on EAE disease course. *Mol Immunol* 2002;38:1373–8.
- [78] Tanzola MB, Robbie-Ryan M, Gutekunst CA, Brown MA. Mast cells exert effects outside the central nervous system to influence experimental allergic encephalomyelitis disease course. *J Immunol* 2003;171:4385–91.
- [79] Sayed BA, Christy AL, Walker ME, Brown MA. Meningeal mast cells affect early T cell central nervous system infiltration and blood–brain barrier integrity through TNF: a role for neutrophil recruitment? *J Immunol* 15-6-2010;184:6891–900.
- [80] Theoharides TC. Mast cells: the immune gate to the brain. *Life Sci* 1990;46:607–17.
- [81] Silver R, Silverman A-J, Vitkovic L, Lederhendler II. Mast cells in the brain: evidence and functional significance. *Trends Neurosci* 1996;19:25–31.
- [82] Robbie-Ryan M, Tanzola MB, Secor VH, Brown MA. Cutting edge: both activating and inhibitory Fc receptors expressed on mast cells regulate experimental allergic encephalomyelitis disease severity. *J Immunol* 15-2-2003;170:1630–4.
- [83] Piconese S, Costanza M, Musio S, Tripodo C, Poliani PL, Gri G, et al. Exacerbated experimental autoimmune encephalomyelitis in mast-cell-deficient Kit<sup>W-sh/W-sh</sup> mice. *Lab Invest* 2011;91:627–41.
- [84] Li H, Nourbakhsh B, Safavi F, Li K, Xu H, Cullimore M, et al. Kit<sup>(W-sh)</sup> mice develop earlier and more severe experimental autoimmune encephalomyelitis due to absence of immune suppression. *J Immunol* 1-7-2011;187:274–82.
- [85] Piliponsky AM, Chen CC, Grimbaldston MA, Burns-Guydish SM, Hardy J, Kalesnikoff J, et al. Mast cell-derived TNF can exacerbate mortality during severe bacterial infections in C57BL/6-Kit<sup>W-sh/W-sh</sup> mice. *Am J Pathol* 2010;176:926–38.

- [86] Kassiotis G, Kranidioti K, Kollias G. Defective CD4T cell priming and resistance to experimental autoimmune encephalomyelitis in TNF-deficient mice due to innate immune hypo-responsiveness. *J Neuroimmunol* 1-10-2001;119:239–47.
- [87] Constantinescu CS, Farooqi N, O'Brien K, Gran B. Experimental autoimmune encephalomyelitis (EAE) as a model for multiple sclerosis (MS). *Br J Pharmacol* 2011;164:1079–106.
- [88] Habib KE, Gold PW, Chrousos GP. Neuroendocrinology of stress. *Endocrinol Metab Clin North Am* 2001;30:695–728.
- [89] Huitinga I, Erkut ZA, van Beurden D, Swaab DF. Impaired hypothalamus–pituitary–adrenal axis activity and more severe multiple sclerosis with hypothalamic lesions. *Ann Neurol* 2004;55:37–45.
- [90] Wei T, Lightman SL. The neuroendocrine axis in patients with multiple sclerosis. *Brain* 1997;120(Pt 6):1067–76.
- [91] Huitinga I, Erkut ZA, van Beurden D, Swaab DF. The hypothalamo-pituitary–adrenal axis in multiple sclerosis. *Ann N Y Acad Sci* 2003;992:118–28.
- [92] Limone P, Ferrero B, Calvelli P, Del Rizzo P, Rota E, Berardi C, et al. Hypothalamic–pituitary–adrenal axis function and cytokine production in multiple sclerosis with or without interferon-beta treatment. *Acta Neurol Scand* 2002;105:372–7.
- [93] Johnson RR, Storts R, Welsh Jr TH, Welsh CJ, Meagher MW. Social stress alters the severity of acute Theiler's virus infection. *J Neuroimmunol* 2004;148:74–85.
- [94] Mohr DC, Pelletier D. A temporal framework for understanding the effects of stressful life events on inflammation in patients with multiple sclerosis. *Brain Behav Immun* 2006;20:27–36.
- [95] Karalis K, Sano H, Redwine J, Listwak S, Wilder RL, Chrousos GP. Autocrine or paracrine inflammatory actions of corticotropin-releasing hormone in vivo. *Science* 1991;254:421–3.
- [96] Chrousos GP. The hypothalamic–pituitary–adrenal axis and immune-mediated inflammation. *N Engl J Med* 1995;332:1351–62.
- [97] Esposito P, Basu S, Letourneau R, Jacobson S, Theoharides TC. Corticotropin-releasing factor (CRF) can directly affect brain microvessel endothelial cells. *Brain Res* 2003;968:192–8.
- [98] Rozniecki JJ, Dimitriadou V, Lambracht-Hall M, Pang X, Theoharides TC. Morphological and functional demonstration of rat dura mast cell–neuron interactions in vitro and in vivo. *Brain Res* 1999;849:1–15.
- [99] Esposito P, Chandler N, Kandere-Grzybowska K, Basu S, Jacobson S, Connolly R, et al. Corticotropin-releasing hormone (CRH) and brain mast cells regulate blood–brain–barrier permeability induced by acute stress. *J Pharmacol Exp Ther* 2002;303:1061–6.
- [100] Donelan J, Boucher W, Papadopoulou N, Lytinas M, Papaliodis D, Theoharides TC. Corticotropin-releasing hormone induces skin vascular permeability through a neurotensin-dependent process. *Proc Natl Acad Sci U S A* 2006;103:7759–64.
- [101] Theoharides TC, Konstantinidou A. Corticotropin-releasing hormone and the blood–brain–barrier. *Front Biosci* 2007;12:1615–28.
- [102] Vasiadi M, Therianou A, Alysandratos KD, Katsarou-Katsari A, Petrakopoulou T, Theoharides A, et al. Serum neurotensin (NT) is increased in psoriasis and NT induces VEGF release from human mast cells. *Br J Dermatol* 27-1-2012;166:1349–52.
- [103] Alysandratos K-D, Asadi S, Angelidou A, Zhang B, Sismanopoulos N, Yang H, et al. Neurotensin and CRH interactions augment human mast cell activation. *PLoS One* 2012;7(11):48934.
- [104] Asadi S, Theoharides TC. Corticotropin-releasing hormone and extracellular mitochondria augment IgE-stimulated human mast-cell vascular endothelial growth factor release, which is inhibited by luteolin. *J Neuroinflammation* 4-5-2012;9:85.
- [105] Esposito P, Gheorghe D, Kandere K, Pang X, Conally R, Jacobson S, et al. Acute stress increases permeability of the blood–brain–barrier through activation of brain mast cells. *Brain Res* 2001;888:117–27.
- [106] Sharma HS, Cervos-Navarro J, Dey PK. Increased blood–brain barrier permeability following acute short-term swimming exercise in conscious normotensive young rats. *Neurosci Res* 1991;10:211–21.
- [107] Benou C, Wang Y, Imitola J, VanVlerken L, Chandras C, Karalis KP, et al. Corticotropin-releasing hormone contributes to the peripheral inflammatory response in experimental autoimmune encephalomyelitis. *J Immunol* 1-5-2005;174:5407–13.
- [108] Sugama S, Takenouchi T, Fujita M, Kitani H, Hashimoto M. Cold stress induced morphological microglial activation and increased IL-1beta expression in astroglial cells in rat brain. *J Neuroimmunol* 2011;233:29–36.
- [109] Sugama S, Fujita M, Hashimoto M, Conti B. Stress induced morphological microglial activation in the rodent brain: involvement of interleukin-18. *Neuroscience* 25-5-2007;146:1388–99.
- [110] Hinwood M, Morandini J, Day TA, Walker FR. Evidence that microglia mediate the neurobiological effects of chronic psychological stress on the medial prefrontal cortex. *Cereb Cortex* 2012;22:1442–54.
- [111] Wang W, Ji P, Dow KE. Corticotropin-releasing hormone induces proliferation and TNF-alpha release in cultured rat microglia via MAP kinase signalling pathways. *J Neurochem* 2003;84:189–95.
- [112] Martin S, Dicu E, Vincent JP, Mazella J. Neurotensin and the neurotensin receptor-3 in microglial cells. *J Neurosci Res* 1-8-2005;81:322–6.
- [113] Singh LK, Pang X, Alexacos N, Letourneau R, Theoharides TC. Acute immobilization stress triggers skin mast cell degranulation via corticotropin-releasing hormone, neurotensin and substance P: a link to neurogenic skin disorders. *Brain Behav Immun* 1999;13:225–39.
- [114] Skaper SD, Giusti P, Facci L. Microglia and mast cells: two tracks on the road to neuroinflammation. *FASEB J* 19-4-2012;26:3103–17.
- [115] Zhang S, Zeng X, Yang H, Hu G, He S. Mast cell tryptase induces microglia activation via protease-activated receptor 2 signaling. *Cell Physiol Biochem* 2012;29:931–40.
- [116] Jeffery DR. Recent advances in treating multiple sclerosis: efficacy, risks and place in therapy. *Ther Adv Chronic Dis* 2013;4:45–51.
- [117] Schrempf W, Ziemssen T. Glatiramer acetate: mechanisms of action in multiple sclerosis. *Autoimmun Rev* 2007;6:469–75.
- [118] Aharoni R. The mechanism of action of glatiramer acetate in multiple sclerosis and beyond. *Autoimmun Rev* 2013;12(5):543–53.
- [119] Pul R, Moharreggh-Khiabani D, Skuljec J, Skripuletz T, Garde N, Voss EV, et al. Glatiramer acetate modulates TNF-alpha and IL-10 secretion in microglia and promotes their phagocytic activity. *J Neuroimmune Pharmacol* 2011;6:381–8.
- [120] Mohr DC, Lovera J, Brown T, Cohen B, Neylan T, Henry R, et al. A randomized trial of stress management for the prevention of new brain lesions in MS. *Neurology* 31-7-2012;79:412–9.
- [121] Artemiadis AK, Vervanioti AA, Alexopoulos EC, Rombos A, Anagnostouli MC, Darviri C. Stress management and multiple sclerosis: a randomized controlled trial. *Arch Clin Neuropsychol* 2012;27:406–16.
- [122] Zoumakis E, Chrousos GP. Corticotropin-releasing hormone receptor antagonists: an update. *Endocr Dev* 2010;17:36–43.
- [123] Zappulla JP, Arock M, Mars LT, Liblau RS. Mast cells: new targets for multiple sclerosis therapy? *J Neuroimmunol* 2002;131:5–20.
- [124] Middleton EJ, Kandaswami C, Theoharides TC. The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease and cancer. *Pharmacol Rev* 2000;52:673–751.
- [125] Rezaei-Zadeh K, Ehrhart J, Bai Y, Sanberg PR, Bickford P, Tan J, et al. Apigenin and luteolin modulate microglial activation via inhibition of STAT1-induced CD40 expression. *J Neuroinflammation* 2008;5:41.
- [126] Jang S, Kelley KW, Johnson RW. Luteolin reduces IL-6 production in microglia by inhibiting JNK phosphorylation and activation of AP-1. *Proc Natl Acad Sci U S A* 27-5-2008;105:7534–9.
- [127] Dirschler K, Karlstetter M, Ebert S, Kraus D, Hlawatsch J, Walczak Y, et al. Luteolin triggers global changes in the microglial transcriptome leading to a unique anti-inflammatory and neuroprotective phenotype. *J Neuroinflammation* 14-1-2010;7:3.
- [128] Kandere-Grzybowska K, Kempuraj D, Cao J, Cetrulo CL, Theoharides TC. Regulation of IL-1-induced selective IL-6 release from human mast cells and inhibition by quercetin. *Br J Pharmacol* 2006;148:208–15.
- [129] Kimata M, Shichijo M, Miura T, Serizawa I, Inagaki N, Nagai H. Effects of luteolin, quercetin and baicalin on immunoglobulin E-mediated mediator release from human cultured mast cells. *Clin Exp Allergy* 2000;30:501–8.
- [130] Kempuraj D, Madhappan B, Christodoulou S, Boucher W, Cao J, Papadopoulou N, et al. Flavonols inhibit proinflammatory mediator release, intracellular calcium ion levels and protein kinase C theta phosphorylation in human mast cells. *Br J Pharmacol* 2005;145:934–44.
- [131] Park HH, Lee S, Son HY, Park SB, Kim MS, Choi EJ, et al. Flavonoids inhibit histamine release and expression of proinflammatory cytokines in mast cells. *Arch Pharm Res* 2008;31:1303–11.
- [132] Hendriks JJ, de Vries HE, van der Pol SM, van den Berg TK, van Tol EA, Dijkstra CD. Flavonoids inhibit myelin phagocytosis by macrophages; a structure–activity relationship study. *Biochem Pharmacol* 2003;65:877–85.
- [133] Aktas O, Prozorovski T, Smorodchenko A, Savaskan NE, Lauster R, Kloetzel PM, et al. Green tea epigallocatechin-3-gallate mediates T cellular NF-kappa B inhibition and exerts neuroprotection in autoimmune encephalomyelitis. *J Immunol* 1-11-2004;173:5794–800.
- [134] Muthian G, Bright JJ. Quercetin, a flavonoid phytoestrogen, ameliorates experimental allergic encephalomyelitis by blocking IL-12 signaling through JAK–STAT pathway in T lymphocyte. *J Clin Immunol* 2004;24:542–52.
- [135] Hendriks JJ, Alblas J, van der Pol SM, van Tol EA, Dijkstra CD, de Vries HE. Flavonoids influence monocyte GTPase activity and are protective in experimental allergic encephalitis. *J Exp Med* 20-12-2004;200:1667–72.
- [136] Kempuraj D, Tagen M, Iliopoulos BP, Clemons A, Vasiadi M, Boucher W, et al. Luteolin inhibits myelin basic protein-induced human mast cell activation and mast cell dependent stimulation of Jurkat T cells. *Br J Pharmacol* 2008;155:1076–84.
- [137] Kang HK, Ecklund D, Liu M, Datta SK. Apigenin, a non-mutagenic dietary flavonoid, suppresses lupus by inhibiting autoantigen presentation for expansion of autoreactive Th1 and Th17 cells. *Arthritis Res Ther* 2009;11:R59.
- [138] Okamoto Y, Tanaka M, Fukui T, Masuzawa T. Brazilian propolis inhibits the differentiation of Th17 cells by inhibition of interleukin-6-induced phosphorylation of signal transducer and activator of transcription 3. *Immunopharmacol Immunotoxicol* 2012;34:803–9.
- [139] Theoharides TC. Luteolin as a therapeutic option for multiple sclerosis. *J Neuroinflammation* 2009;6:29.
- [140] Sternberg Z, Chadha K, Lieberman A, Drake A, Hojnacki D, Weinstock-Guttman B, et al. Immunomodulatory responses of peripheral blood mononuclear cells from multiple sclerosis patients upon in vitro incubation with the flavonoid luteolin: additive effects of IFN-beta. *J Neuroinflammation* 2009;6:28.
- [141] Steelman AJ, Dean DD, Young CR, Smith III R, Prentice TW, Meagher MW, et al. Restraint stress modulates virus specific adaptive immunity during acute Theiler's virus infection. *Brain Behav Immun* 2009;23:830–43.